

ORIGINAL ARTICLE**Protective Effect of Methanolic Extract of *Tamarindus Indica* Leaves in Ethanol-Induced Esophageal Ulcer in Adult Wistar Rats**Umar S. R.¹, Isyaku M. U.^{1,2}, Mwabaleke J. A.², Etukudo E.M.^{2,*}¹Department of Anatomy, Faculty of Basic Medical Sciences, Yusuf Maitama Sule University, Kano, Nigeria²Department of Human Anatomy, Faculty of Biomedical Sciences, Kampala International University, Western Campus, Uganda**ABSTRACT**

Tamarind (*Tamarindus indica L.*) possess anti-ulceration, wound healing, anti-inflammatory and antipyretic properties. This makes it useful in the treatment of various clinical challenges including gastrointestinal tract diseases. The aim of this study was to determine the protective effect of methanolic extract of *Tamarindus indica* leaves on esophageal ulcers in adult Wistar rats. Twelve rats (12) with average body weights of 130 ± 20 grams (g) were divided into four groups (A, B, C and D) (n=4). Group A (control) received 0.5 ml/kg body weight (bw) of distilled water, Group B received 0.5 ml/kg bw of 80% ethanol, Group C (Low dose) received 200 mg/kg of methanolic extract of *Tamarindus indica* leaves + 0.5ml/kg of 80% Ethanol and Group D (High dose) received 300 mg/kg of methanolic extract of *Tamarindus indica* leaves + 0.5ml/kg of 80% Ethanol. All administrations were done via gastric intubation for 6 days. At the end of the experimental period sections of the esophagus were collected for histological studies. The group administered 0.5ml/kg of 80% ethanol when compared with the control group, showed ulceration along the surface margin of mucosal layers and damage to submucosal glands. The group administered high and low doses of methanolic extract of *Tamarindus indica* leaves + 0.5ml/kg of 80% ethanol when compared with the control group, revealed an intact histoarchitecture. METL was protective in the ethanol induced esophageal ulceration.

Keywords: Esophageal ulcer, *Tamarindus indica*, Ethanol.***Corresponding Author**

Etukudo, Ekom Monday; Department of Human Anatomy, Faculty of Biomedical Sciences, Kampala International University, Western Campus, Uganda; ekometukudo@gmail.com, +2348068792326

Citing this articleUmar S. R., Isyaku M. U., Mwabaleke J. A., Etukudo E.M. Protective Effect of Methanolic Extract of *Tamarindus Indica* Leaves in Ethanol-Induced Esophageal Ulcer in Adult Wistar Rats. KIU J. Health Sci, 2023; 3(2);

Conflict of Interest: The authors declare no conflict of interest.

INTRODUCTION

Ulcers are a common gastrointestinal condition that can cause discomfort and bleeding (1,2). It is essentially an inflammatory breach in the skin or the mucus membrane lining the gastrointestinal system (3). The ulceration takes place when a healthy equilibrium is disrupted due to increased aggressiveness or decreased mucosal resilience. It might be related to habitual drug use, irregular eating habits, stress, or other risk factors. Peptic ulcers is a wide term that covers ulcers affecting the digestive system in the stomach or duodenum (4,5). There are several forms of ulcers such as mouth ulcers, esophageal ulcers, peptic ulcers, and genital ulcers, however esophageal ulcers are one of the most prevalent (affecting about 10% of the World's population) and is rare cause of upper gastrointestinal bleeding (6).

Ethanol consumption have been linked with various health problems including ulcers and neurological disturbances (7–9)

Plants have been used as traditional remedies and pharmacopeial drug from ancient times, to reduce human suffering (10,11). *Tamarindus indica* (Tamarind, English; Icheku, Igbo; Awin, Yoruba; Tsamiya-Kurm, Hausa; Chwa, Acholi; Enkooge, Luganda; and Apeduru, Itesot) (12,13), is one of the well-cultivated and familiar plants all over the world which serves as a medicinal plant for the treatment of several diseases and serves as a nutritional plant for malnutrition. Tamarind is a huge broad-leaved, tropical tree found in regions of India and Asia (14). *Tamarindus indica* has highly essential phytochemicals including phenolic compounds, uronic acid, tartaric acid, malic acid, pectin and glucose (14,15). It contains vital elements including copper, calcium, iron, manganese, sodium, magnesium, phosphorus, potassium and zinc (16,17). *Tamarindus indica* has been utilized in Ayurveda to treat gastrointestinal, inflammatory, neurological, and digestive issues (18,19). However, there is paucity of literature on the histological changes in the esophagus following alcohol induced ulceration. More so, previous study on the effect *Tamarindus indica* on ulceration focused only on the

peptic ulcer. This study aimed to assess the histoprotective ability of a methanolic extract of *Tamarindus indica* leaves (METL) in ethanol-induced esophageal ulcers.

MATERIALS AND METHODS

Ethical Approval

The study was authorized by the Faculty of Basic Medical Sciences Research Committee and registered as UG14ANT010.

Plant Collection and Preparation

The Tamarind leaves were collected fresh from Dawakin Tofa Local Government, along Dawakin Tofa Science College, Kano State. The plant leaves were identified at Department of Plant Biology in Bayero University, Kano with identification number BUK/HAN/0074. They were then dried for 7 days under the shade and pulverized using an electric grinder (Nima Brand; SKU:NI467HA083PYFNAFAMZ, Jumia). Methanolic extraction of the plant was done using Soxhlet Apparatus as outlined by Livingston (14).

Animal Procurement

Twelve (12) Wistar rats (both male and female) with average body weights of 130 ± 20 g, were used for the study and obtained from the department of Biological Sciences, Bayero University Kano and were acclimatized for one week (7 days) in the Department of Anatomy, Yusuf Maitama Sule University Kano (YMSK). The rats were housed in colony cages at convenient temperature and relative humidity. The rats had unrestricted access to normal pelletized grower feed and drinking water. Animals were handled in accordance with laboratory animal care and use protocols, as well as ethical principles for experimental animal research. The study was approved by the Department of Human YMSK research committee.

Experimental Design

After 7 days of acclimatization, the twelve (12) Wistar rats had their weights taken and divided into four (4) subgroups of three rats per cage.

The LD₅₀ for methanolic extract of *Tamarindus indica* leaves (METL) was ≥ 5000 as adopted from Highab *et al.* (20). 1/25 fraction of the LD₅₀ value of METL was employed to obtain 200 mg/kg dosage (as low dose) while 100 mg/kg was added to make up the high dose.

Group A (control) received 0.5 ml/kg body weight (bw) of distilled water, Group B received 0.5 ml/kg bw of 80% ethanol, Group C (Low dose) received 200mg/kg of methanolic extract of *Tamarindus indica* leaves (METL) + 0.5 ml/kg of 80% Ethanol and Group D (High dose) received 300 mg/kg of METL + 0.5 ml/kg of 80% Ethanol. The various dosages for administration and protocol were adopted from Kumar *et al.*, [15]. All administrations were done via gastric intubation; methanolic extract of *Tamarindus indica* leaves (METL) was administered for 6 days to rats in the treatment groups (Group B, C & D), followed by ethanol administration on the last day. Prior to the administration of ethanol, the animals were starved and dehydrated. The animals being studied were sacrificed 45 minutes after being administered ethanol. The esophagus was dissected out, and washed with normal saline (0.9%), and the inner surfaces were examined for ulcerations [15].

Tissue Collection and Processing

The experimental animals were humanely sacrificed following chloroform inhalation. Each of the rats was fixed on the dissecting board with its ventral surface facing upward. An incision was made from the lower jaw up to the anal orifice of the rats revealing the abdominal contents. The esophagus together with the stomach was located and harvested. The samples were promptly fixed with 10% neutral buffered formalin. The tissues were grossed and prepared for regular histological analysis. Sections of the esophagus tissues were stained with Hematoxylin and Eosin (H and E) (21). The microscopic images were obtained with a light microscope (Olympus BH2) and a Nikon Digital Sight DS-L1 (Nikon Corporation, Japan).

RESULTS

The result from the histological examination of the lower part of the esophagus in the control group revealed normal histo-architecture with an intact layer (with normal mucosa, submucosa, muscularis mucosa, and adventitia/Serosa) (Figure 1). The administered 0.5ml/kg body weight of 80% ethanol showed ulceration along the surface margin of mucosal layers and damage of submucosal glands (Figure 2). The groups administered 200 mg/kg and 300 mg/kg body weight of the extract followed by 0.5ml/kg of 80% ethanol respectively revealed intact layers (Figure 3 and Figure 4).

DISCUSSION

Ulceration occurs when the natural balance is broken due to a rise in aggression or diminished mucosal resilience. It could be brought on by consistent drug use, strange eating patterns, stress, and other factors. There are numerous different forms of ulcers, including vaginal, esophageal, peptic, and oral ulcers. Because of this, esophageal ulcers are among the more prevalent forms of ulcers and an uncommon source of upper gastrointestinal hemorrhage (1,2).

The present study determined the protective effect of METL on esophageal ulcers in adult Wistar rats. The result of the present study revealed that the administration of METL was protective in ethanol-induced esophageal ulceration. This observation is in line with the finding of Kumar *et al.* (22) who discovered that *T. indica* seed extract had a dose-dependent protective effect against peptic ulcer models produced by ibuprofen, alcohol, and pylorus closure. Improvement in the histoarchitecture of the esophagus in the present can be hinged on the rich phytochemical composition of *Tamarindus indica* (23). Plants rich in phytochemicals such as polyphenols, flavonoids, and tannin just like *Tamarindus indica* are known to possess protective or therapeutic potential (24–27). The antioxidant activities of phytochemicals like polyphenols and flavonoids play a protective role against reactive oxygen species; hence, preserving the cellular integrity of the esophagus (23–25). Tannin present in *Tamarindus indica* interferes with the development of ulcers by causing protein accumulation and vasoconstriction (5,22,28,29). The current study's findings, however simple, provide insight into

Tamarindus indica's protective capability during prenatal ethanol exposure.

CONCLUSION

The present study demonstrated that the methanolic extract of leaves of tamarind minimizes the histopathological changes in the esophagus induced by alcohol in adult Wistar rats. Thus, the extract possesses anti-ulcer properties and may have a potential therapeutic agent for ulceration by alcohol.

REFERENCES

- Ananthkrishnan AN, Xavier RJ. Gastrointestinal Diseases. Hunter's Trop Med Emerg Infect Dis [Internet]. 2020 Jan 1 [cited 2024 Jan 15];16–26. Available from: <https://linkinghub.elsevier.com/retrieve/pii/B978032355512800003X>
- Costable NJ, Greenwald DA. Upper Gastrointestinal Bleeding. Geriatr Gastroenterol Second Ed [Internet]. 2021 Jan 1 [cited 2024 Jan 15];1289–304. Available from: https://link.springer.com/referenceworkentry/10.1007/978-3-030-30192-7_47
- Jaiswal, Wal AK, Wal P, Singh AP. PEPTIC ULCER: A REVIEW ON ETIOLOGY, PATHOGENESIS AND TREATMENT. Pathog Treat Asian J Pharm Educ Res [Internet]. 2021 [cited 2024 Jan 15];10(4):1–17. Available from: <https://dx.doi.org/10.38164/AJPER/9.4.2021.01-17>
- Awuchi CG, Saha P, Amle VS, Nyarko RO, Kumar R, Boateng EA, et al. A Study of Various Medicinal Plants used in Ulcer Treatment: A Review. J Res Appl Sci Biotechnol [Internet]. 2023 Mar 15 [cited 2024 Jan 15];2(1):234–46. Available from: <https://www.jrasb.com/index.php/jrasb/article/view/167>
- P. S. Sreeja, K. Arunachalam and PT. Evaluation of Anti-ulcer Potential of *Sphenodesme involucreta* var. *paniculata* (C.B. Clarke) Munir Leaves on Various Gastric Aggressive Factors [Internet]. Phytomedicine: Research and Development. 2020 [cited 2024 Jan 15]. Available from: https://ebrary.net/158639/health/evaluation_anti_ulcer_potential_sphenodesme_involucreta_paniculata_clarke_munir_leaves_various
- Srivastav Y, Kumar V, Srivastava Y, Kumar M. Peptic Ulcer Disease (PUD), Diagnosis, and Current Medication-Based Management Options: Schematic Overview. J Adv Med Pharm Sci. 2023 Dec 2;25(11):14–27.
- Usman IM, Iliya IA, Ivang AE, Ssempijja F, Ojewale AO, Yusuf HR. Microanatomical and biochemical changes of the cerebellum following ethanol gavage in adult Wistar rats. Anat J Africa [Internet]. 2019 Sep 13 [cited 2024 Jan 17];8(2):1662–9. Available from: <https://www.ajol.info/index.php/aja/article/view/189708>
- Usman I, Buraimoh A, Ibegbu A. Histological and biochemical studies of *Tamarindus indica* pulp extract on the cerebral cortex in prenatal ethanol exposure in Wistar rats. J Exp Clin Anat [Internet]. 2016 Jul 1 [cited 2023 Nov 20];15(2):96–96. Available from: <https://go.gale.com/ps/i.do?p=AONE&sw=w&issn=15962393&v=2.1&it=r&id=GALE%7CA483768257&sid=googleScholar&linkaccess=fulltext>
- Sheikh M, Roshandel G, McCormack V, Malekzadeh R. Current Status and Future Prospects for Esophageal Cancer. Cancers 2023, Vol 15, Page 765 [Internet]. 2023 Jan 26 [cited 2024 Jan 17];15(3):765. Available from: <https://www.mdpi.com/2072-6694/15/3/765/htm>
- Qadir SU, Raja V. Herbal medicine: Old practice and modern perspectives. Phytomedicine A Treasure Pharmacol Act Prod from Plants. 2021 Jan 1;149–80.
- Ayuba JT, Okesina AA, Usman IM, Ajenikoko MK, Pius T, Kusiima N, et al. *Galinsoga parviflora* restored associated

- motor coordination through increased linear distribution of Purkinje Cells in mercury chloride-induced toxicity of mices cerebellum. *African J Cell Pathol.* 2022 Jul;14(1):1–8.
12. Toungos MD. Tamarind (*Tamarindus indicus* L) Fruit of Potential Value But Underutilized in Nigeria. *Int J Innov Food* [Internet]. 2019 [cited 2024 Jan 15];7(1):1–10. Available from: www.seahipaj.org
 13. Okello J, Okullo JBL, Eilu G, Nyeko P, Obua J. Morphological Variations in *Tamarindus indica* LINN. Fruits and Seed Traits in the Different Agroecological Zones of Uganda. *Int J Ecol.* 2018;2018.
 14. Livingston Raja NR, Jegan N, Wesley J. *Pharmacologyonline* 3: 85-92 (2008) Livingston Raja et al. 85 ANTIULCEROGENIC ACTIVITY OF ALCOHOLIC EXTRACT OF THE LEAVES OF TAMARINDUS INDICA (L) ON EXPERIMENTAL ULCER MODELS.
 15. Balkrishna A, Bhadoria K, Mishra S, Kumar A, Singh S, Srivastava D, et al. Ethnomedicinal plants used by Juang and Munda Tribals of Eastern India against gastrointestinal disorders: A narrative review. *Med Plants.* 2023 Jun 1;15(2):245–60.
 16. Yeasmin, Singh. MINERALS AND HEAVY METALS CONTENT STATUS IN SOME FRUITS IN INDIA. *Int Contemp J Sci Educ Technol* [Internet]. 2022 Sep 1 [cited 2024 Jan 15];1:155–72. Available from: <https://journals.iapaar.com/index.php/AAJSET/article/view/79>
 17. Mukherjee DPS. Tamarind (*Tamarindus indica*) Seeds in Health and Nutrition. *Nuts Seeds Heal Dis Prev.* 2020 Jan 1;1:171–82.
 18. Pramila G, Jirekar D, Dhonde Alias Babaji Mahavidyalaya Kada Dist Beed A. *Tamarindus Indica: An Important Medicinal Plants.* 2021;
 19. Usman IM, Agbon AN, Ivang AE, Peter AB, Afodun AM, Okesina AA, et al. Ethyl acetate fraction of *Tamarindus indica* leaf ameliorates aluminium chloride induced neural damage in neonatal Wistar rats. *J Trace Elem Miner.* 2023 Mar 1;3:100047.
 20. Highab SM, Raji I, Abubakar A, Hassan FI. Analgesic Activity of Methanol Leaf Extract of *Tamarindus Indica* Plant in Mice and Rats. *Dutse J Pure Appl Sci.* 2022 Jan 27;7(3b):143–54.
 21. Fischer AH, Jacobson KA, Rose J, Zeller R. Hematoxylin and Eosin Staining of Tissue and Cell Sections. *Cold Spring Harb Protoc* [Internet]. 2008 May 1 [cited 2024 Feb 6];2008(5):pdb.prot4986. Available from: <http://cshprotocols.cshlp.org/content/2008/5/pdb.prot4986.full>
 22. Kumar S, Suman, Sharma S, Kalra P. Antiulcer effect of the methanolic extract of *Tamarindus indica* seeds in different experimental models. *J Pharm Bioallied Sci* [Internet]. 2011 Apr [cited 2024 Jan 15];3(2):236–41. Available from: https://journals.lww.com/jpbs/fulltext/2011/03020/antiulcer_effect_of_the_methanolic_extract_of.9.aspx
 23. Kuru P. *Tamarindus indica* and its health related effects. *Asian Pac J Trop Biomed.* 2014 Sep 1;4(9):676–81.
 24. Devi B, Boruah T. Tamarind (*Tamarindus indica*). Antioxidants Fruits Prop Heal Benefits [Internet]. 2020 Jan 1 [cited 2024 Jan 15];317–32. Available from: https://link.springer.com/chapter/10.1007/978-981-15-7285-2_16
 25. Borquaye LS, Doetse MS, Ofori Baah S, Mensah JA, Sheringham Borquaye L, Baah SO. Anti-inflammatory and anti-oxidant activities of ethanolic extracts of *Tamarindus indica* L. (Fabaceae). *Cogent Chem* [Internet]. 2020 Jan 1 [cited 2024 Jan 15];6(1):1743403. Available from: <https://www.tandfonline.com/doi/abs/10.1080/23312009.2020.1743403>
 26. Garba Y, Bello NU, Bashir M, Mohammed

- SY. In vitro and in vivo analysis of the antioxidant potentials of methanolic extract of *Tamarindus indica* leaves. *Niger J Biochem Mol Biol* [Internet]. 2023 Nov 27 [cited 2024 Jan 15];38(3):104–11. Available from: <https://www.ajol.info/index.php/njbmb/article/view/259372>
27. Sookyng S, Duangjai A, Saokaew S, Phisalprapa P. Botanical aspects, phytochemicals, and toxicity of *Tamarindus indica* leaf and a systematic review of antioxidant capacities of *T. indica* leaf extracts. *Front Nutr* [Internet]. 2022 Sep 20 [cited 2023 Nov 21];9. Available from: [/pmc/articles/PMC9530316/](https://www.frontiersin.org/articles/10.3389/fnut.2022.9530316)
28. Yazarlu O, Iranshahi M, Kashani HRK, Reshadat S, Habtemariam S, Iranshahi M, et al. Perspective on the application of medicinal plants and natural products in wound healing: A mechanistic review. *Pharmacol Res*. 2021 Dec 1;174:105841.
29. Laware R, Bharskar G, Chavan S, Borudeq G. A Review on Polyherbal Antiulcer Formulations. *Res J Pharmacogn Phytochem*. 2022 Apr 28;89–97.

FIGURES

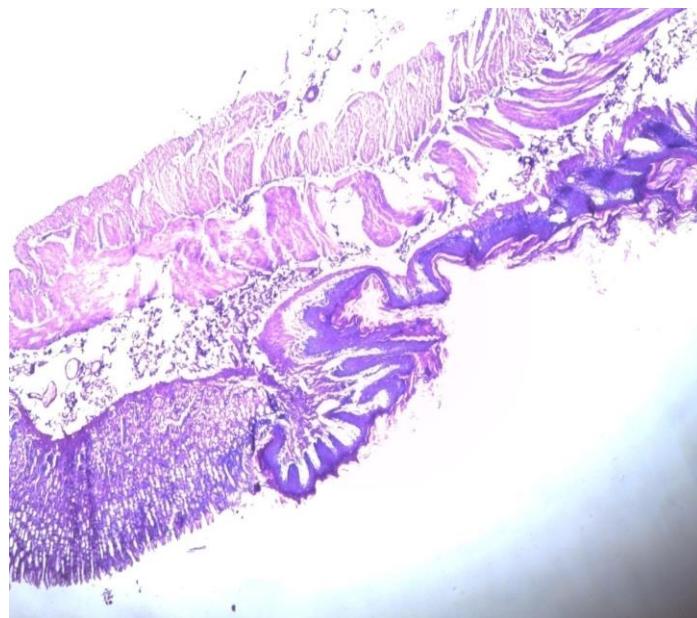


Figure 1: Photomicrograph of esophageal section from the group administered 0.5ml/kg of distilled water (H&E; X40).

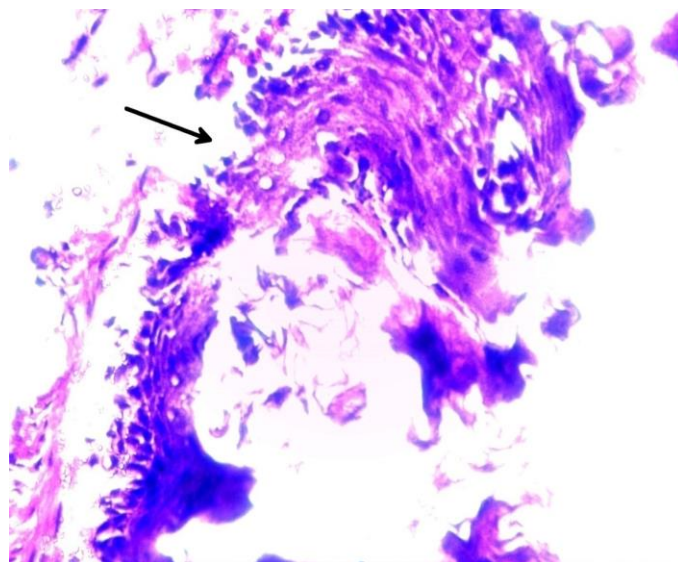


Figure 2: Photomicrograph of esophageal section from the group administered 0.5ml/kg of 80% ethanol (H&E; X40).

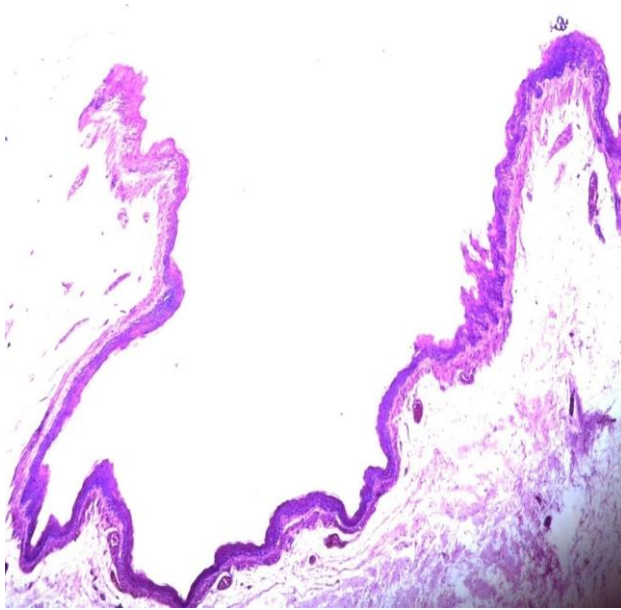


Figure 3: Photomicrograph of esophageal section from the group administered 200mg/kg of extract and 0.5ml/kg of distilled water (H&E; X40).

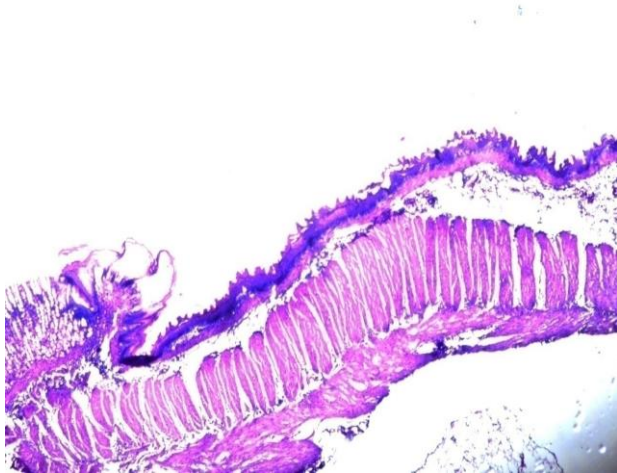


Figure 4: Photomicrograph of esophageal section from the group administered 300mg/kg of extract and 0.5ml/kg of distilled water (H&E; X40).